



## **International Journal of Medical and Pharmaceutical Case Reports**

**12(3): 1-6, 2019; Article no.IJMPCR.50866**  
**ISSN: 2394-109X, NLM ID: 101648033**

# **Detection of Metastatic Breast Carcinoma Cells in Bone Marrow by Flow Cytometry**

**Manoela Lira Reis<sup>1</sup>, Daniella Serafin Couto Vieira<sup>2</sup>, Laura Otto Walter<sup>3</sup>  
and Maria Claudia Santos-Silva<sup>4\*</sup>**

<sup>1</sup>*Polydoro Ernani de São Thiago University Hospital, Federal University of Santa Catarina, Florianópolis, Brazil.*

<sup>2</sup>*Department of Clinical Analysis, Federal University of Santa Catarina, Florianópolis, Brazil.*

<sup>3</sup>*Department of Clinical Pathology, Federal University of Santa Catarina, Florianópolis, Brazil.*

<sup>4</sup>*Department of Pharmacy, Federal University of Santa Catarina, Florianópolis, Brazil.*

### **Authors' contributions**

*This work was carried out in collaboration among all authors. Author MLR designed the study and wrote the first draft of the manuscript. Authors DSCV and LOW managed the analyses of the study. Author MCSS managed the literature searches. All authors read and approved the final manuscript.*

### **Article Information**

DOI: 10.9734/IJMPCR/2019/v12i330105

#### Editor(s):

(1) Dr. Erich Cosmi, Director of Maternal and Fetal Medicine Unit, Department of Woman and Child Health, University of Padua School of Medicine, Padua, Italy.

#### Reviewers:

(1) Michael Bordonaro, Geisinger Commonwealth School of Medicine, USA.

(2) Alessandro Poggi, IRCCS Ospedale Policlinico San Martino, Italy.

Complete Peer review History: <http://www.sdiarticle3.com/review-history/50866>

**Case Study**

**Received 10 June 2019**  
**Accepted 22 August 2019**  
**Published 09 September 2019**

## **ABSTRACT**

Breast cancer is the most common cause of cancer death in women worldwide. Cytological, histological, and immunohistochemical techniques are routine laboratory tests for determining tumor subtypes. Over the past few years, laboratory diagnostic tests for breast cancer have become more complex, sophisticated, and specialized. This report describes the case of a young patient with metastatic breast cancer whose diagnosis was based on flow cytometric analysis of bone marrow aspirate. Flow cytometry showed to be an important tool in cancer diagnosis. Its application as a routine laboratory test for the diagnosis of solid tumors, such as breast cancer, can help provide fast results while increasing diagnostic coverage.

*Keywords: Breast carcinoma; bone marrow; flow cytometry; breast cancer.*

\*Corresponding author: E-mail: [santosdasilvamc@gmail.com](mailto:santosdasilvamc@gmail.com);

## 1. INTRODUCTION

Breast cancer is the most common cause of cancer death in women worldwide [1]. Despite advances in detection strategies and multi-professional approaches, many women are still diagnosed with advanced-stage breast cancer, which decreases their chances of cure, especially in cases of metastasis [2]. Time of detection and histological type are important prognostic factors. Cytological, histological, and immunohistochemical techniques are routine laboratory tests widely used for determining tumor subtypes [2]. Over the years, laboratory diagnostic tests for this cancer have become more complex, sophisticated, and specialized, resulting in faster results and more personalized treatments for each tumor subtype [3].

This report describes the case of a young patient with a history of chronic bone pain. Diagnosis of metastatic breast cancer was based on analysis of bone marrow aspirate by flow cytometry.

## 2. PRESENTATION OF CASE

The patient is a 28-year-old woman, adopted, mother of three children, with a history of postpartum depression, undergoing treatment for lactation mastitis in the right breast. The patient presented with five months of worsening bilateral lumbar pain radiating to the thorax and lower limbs. Two months later, she returned to the hospital with epistaxis, alopecia, lymphadenopathy, exertional dyspnea, petechiae in the lower limbs, and weight loss of 20 kg. Laboratory examination revealed bicytopenia. The patient was admitted to the hospital with fever and night sweats. Clinical findings and patient history favored initial hypothesis of lymphoproliferative neoplasm. Bone marrow

aspirate and biopsy were collected for immunophenotypic, histological, and immunohistochemical examination.

A standardized panel of monoclonal antibodies for samples suspected of hematological neoplasms (anti-CD45-V500, anti-CD34-PerCP-Cy5.5, anti-CD3-APC, anti-CD19-PE-Cy7, anti-CD56-PE, anti-CD38-APC, anti-CD20-PB, and anti-CD8-FITC) was used for flow cytometry. Samples were also labeled with an anti-HER2-PE antibody using a protocol standardized for samples from female patients with suspected non-hematologic malignancies of unknown primary. Sample acquisition was performed on a FACSCanto II (BD Biosciences, San Jose, CA, USA), and data were analyzed using Infinicyt® version 1.7 (Cytognos, Spain).

Cytology of MGG-stained bone marrow aspirate from patient of this case report revealed non-hematological cells with atypical, enlarged nuclei (Fig. 1A to C).

Morphological and flow cytometric analysis of bone marrow aspirate negated the initial hypothesis of lymphoproliferative neoplasm. In the following step, diagnosis of malignant breast neoplasm was confirmed by detection of HER2<sup>+</sup> cells by flow cytometry (Fig. 2A to F). Flow cytometry results; therefore, helped define the antibody panel for immunohistochemistry analysis of bone marrow biopsy.

Immunohistochemistry results (Fig. 3A to 3F) showed the presence of epithelial cells in bone marrow biopsy with positive expression of ER, HER2, GCDPF-15, mammaglobin, the pool of CK, CK7, E-cadherin, and PR. These phenotypic characteristics were compatible with metastatic breast carcinoma in bone marrow.

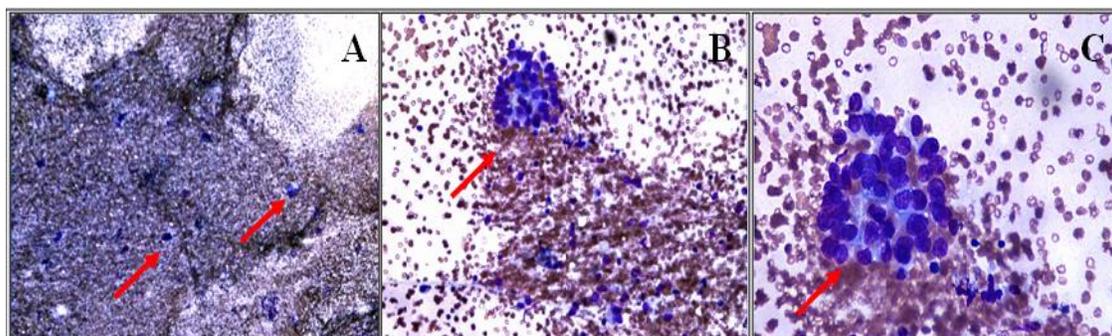
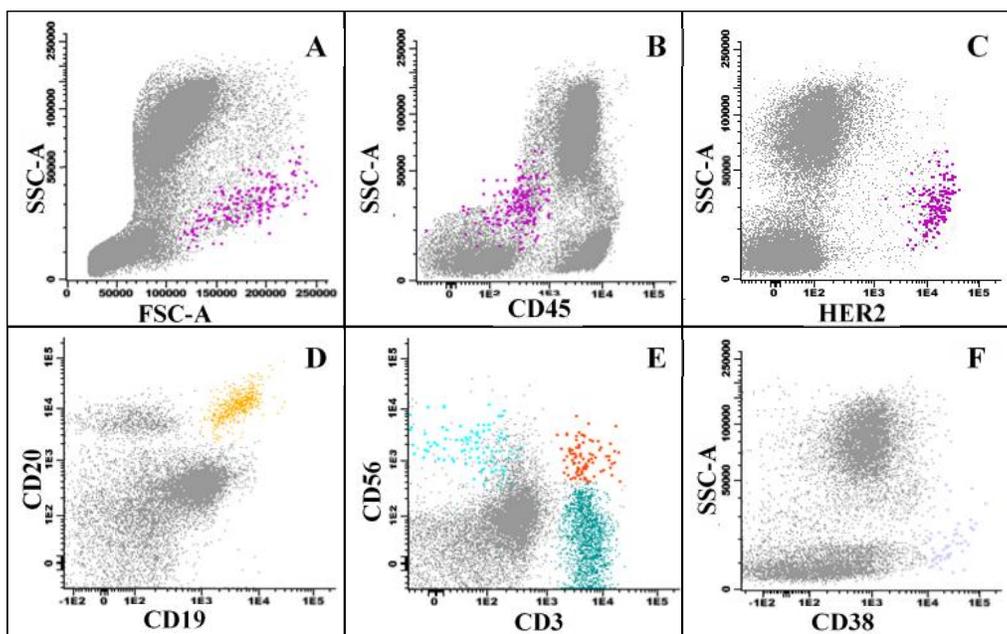
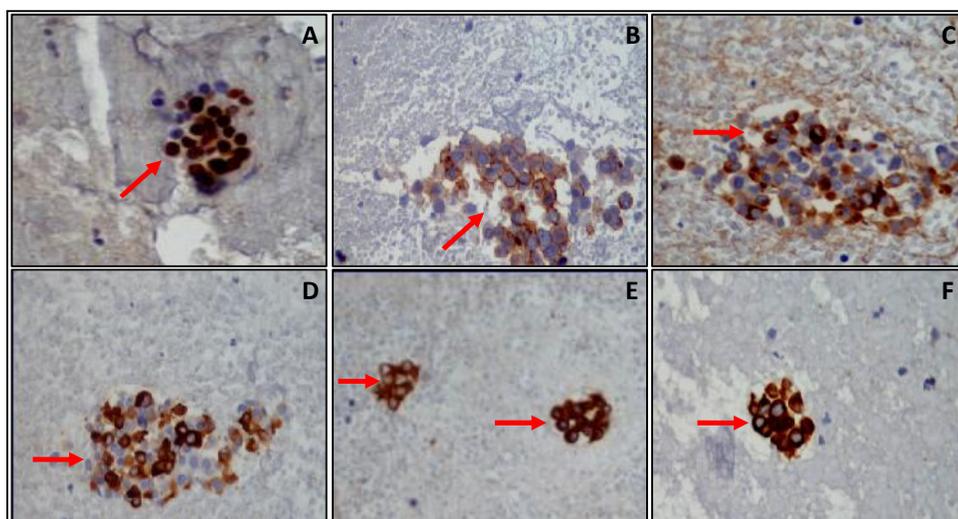


Fig. 1. Myelogram of bone marrow aspirate stained with May-Grünwald-Giemsa at 4× (A), 10× (B), and 40× magnification (C)



**Fig. 2.** Immunophenotypic profile of bone marrow aspirate cells by flow cytometry. (A–C) Size (FSC) and granularity (SSC) of non-hematopoietic cells (CD45<sup>-</sup>/HER2<sup>+</sup>), shown in pink. (D) B lymphocytes (CD19<sup>+</sup>/CD20<sup>+</sup>) highlighted in yellow. (E) T lymphocytes (CD3<sup>+</sup>) highlighted in green, NK cells (CD56<sup>+</sup>) in blue, and NKT cells (CD56<sup>+</sup>/CD3<sup>+</sup>) in orange. (F) Plasma cells (CD38<sup>++</sup>) highlighted in lilac

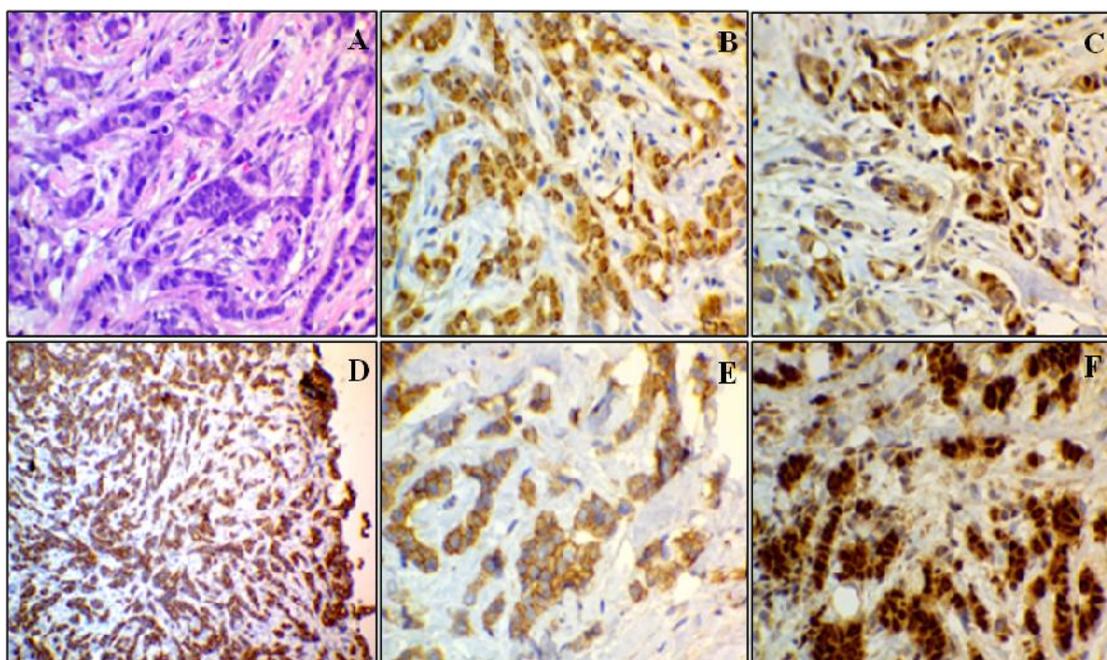


**Fig. 3.** Immunohistochemistry of bone marrow biopsy of the reported case showing presence of epithelial cells with positive reaction to anti-estrogen receptor (A), anti-HER2 (B), anti-GCDFP15 (C), anti-mammaglobin (D), anti-CK pool (E), and anti-CK7 (F) at 100× magnification

Subsequent physical examination revealed a nodule in the right breast, which was biopsied and analyzed. H&E-stained microscopic images showed invasive mammary carcinoma. Immunohistochemistry was carried out using a prognostic panel for breast cancer (ER, PR,

HER2, and Ki67) as well as CK7 and GATA3 antibodies, and positivity for CK7 and GATA3 was observed Fig. 4A to 4F).

The results confirmed the presence of primary breast tumor. Late diagnosis contributed to



**Fig. 4. Morphological and immunohistochemical evaluation of breast biopsy specimen of the reported case (A) Breast biopsy specimen stained with hematoxylin and eosin showing features characteristic of invasive mammary carcinoma ( $\times 100$ ). (B–F) Immunohistochemistry of breast biopsy specimens at  $100\times$  magnification. (B) Breast tissue with positive expression of progesterone receptor (+/Allred 6). (C) Breast tissue with positive expression of estrogen receptor (+/Allred 8). (D) Mammary gland tissue with positive expression of CK7 (+/diffuse). (E) Breast tissue with positive expression of HER2 (+/indeterminate). (F) Breast tissue with positive expression of GATA3 (+/diffuse)**

disease progression and a poor outcome. The patient died six months after diagnosis.

### 3. DISCUSSION

Here we report the case of a young patient diagnosed with metastatic breast carcinoma in bone marrow who died six months after diagnosis. Aggressive phenotypes of neoplasms commonly occur in young women, aged less than 45 years, resulting in poor prognosis and high risk of death. The Brazilian government, aiming to reduce breast cancer-related mortality rates, recommends screening from 40 years of age onward for women in the low-risk group and 30 years onward for women in the high-risk group [4]. The level of risk is determined by family history, environmental factors, parity, and lifestyle habits [5,6].

In the case of the patient presented here, her young age, unknown family history, and the overall clinical picture made the diagnosis difficult. The initial suspicion of lymphoma misled

the multidisciplinary team. Flow cytometry helped the team achieve quick results, despite the advanced stage of the disease. This scenario emphasizes the importance of laboratory techniques that assist in rapidly establishing the correct diagnosis. We highlight that the use of anti-HER2 antibody for the differential diagnosis of lymphoma by flow cytometry was an important step in reaching a final diagnosis. Detection of HER2<sup>+</sup> expression in non-hematopoietic cells in bone marrow alerted to the presence of metastasis, suggesting malignant breast neoplasm, the most common neoplasm in women [7,8].

Flow cytometric immunophenotyping is a relevant method that has made great contributions to the diagnosis of hematological malignancies [9,10]. The technique is able to phenotypically characterize and differentiate abnormal cell populations from normal populations, even at low concentrations, by means of antigen–antibody reactions. Reliable results are obtained in less than 4 h of

processing [11]. Although this method is widely used as a routine diagnostic tool for hematological disorders, its application in solid tumor diagnosis remains limited, to a large extent, to research purposes. [12,13,14]. Thus cases reports should be describe showing the importance of the flow cytometry to diagnosis of solid tumors.

#### 4. CONCLUSION

The results presented in this case report show the importance of flow cytometry in the laboratory routine. In this case, the initial diagnostic hypothesizes lymphoproliferative neoplasia, however the inclusion of the HER2 immunostaining on the screening panel by flow cytometric was essential for directing the diagnosis of the of metastatic breast carcinoma in the bone marrow. Thus, the flow cytometry was an essential tool for the quick and rapid conclusion of this case. The results, obtained by flow cytometry, guided the markers of breast carcinoma investigation by immunohistochemistry in early bone marrow biopsy and allowed the correct clinical management of the case. In this sense, the flow cytometry showed to be an important tool in cancer diagnosis. Its application, as a routine laboratory test for the diagnosis of solid tumors, can help provide fast results while increasing diagnostic coverage.

#### CONSENT

All authors declare that 'written informed consent was obtained from the patient for publication of this paper and accompanying images.

#### ETHICAL APPROVAL

The patient agreed to participate in this study and signed an informed consent form approved by the Human Research Ethics Committee of the Federal University of Santa Catarina, Brazil - CEPISH/USFC no. 1.691.983/2016 - (Supplementary File).

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer

- statistics: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68(6):394-424.
2. Winters S, Martin C, Murphy D, Shokar NK. Breast cancer epidemiology, prevention, and screening. *Prog. Mol. Biol. Transl Sci.* 2017;151:1-32.
3. Jafari SH, Saadatpour Z, Salmaninejad A, Momeni F, Mokhtari M, Nahand JS, Rahmati M, Mirzaei H, Kianmehr M. Breast cancer diagnosis: Imaging techniques and biochemical markers. *J. Cell Physiol.* 2018;233(7):5200-5213.
4. Migowski A, Dias MBK, Nadanovsky P, Silva GA, Sant'Ana DR, Stein AT. Guidelines for early detection of breast cancer in Brazil. III – Challenges for implementation. *Cad Saúde Pública.* 2018; 34(6).
5. Klarenbach S, Sims-Jones N, Lewin G, Singh H, Thériault G, Tonelli M, Doull M, Courage S, Garcia AJ, Thombs BD. Recommendations on screening for breast cancer in women aged 40–74 years who are not at increased risk for breast cancer. *CMAJ.* 2018;10;190(49).
6. Rojas K, Stuckey A. Breast cancer epidemiology and risk factors. *Clin. Obstet Gynecol.* 2016;59(4):651-672.
7. Li X, Zhang Y, Meisel J, Jiang R, Behera M, Peng L. Validation of the newly proposed American Joint Committee on Cancer (AJCC) breast cancer prognostic staging group and proposing a new staging system using the National Cancer Database. *Breast Cancer Res Treat.* 2018; 171(2):303-313.
8. Wang D, Xu J, Shi G, Yin G. Molecular markers' progress of breast cancer treatment efficacy. *J. Cancer Res Ther.* 2015;11(Suppl 1):C11-5.
9. Swerdlow SH, Campo E, Harris NL, Jasse, E, Pireli SA, Stein H, Thiele J, Vardiman, JW. WHO classification of tumours of hermatopoietic and lymphoid tissues. 4<sup>th</sup> Ed. Lyon: IARC; 2017.
10. Betters DM. Use of flow cytometry in clinical practice. *J. Adv. Pract. Oncol.* 2015;6:435–440.
11. Adan A, Alizada G, Kiraz Y, Baran Y, Nalbant A. Flow cytometry: Basic principles and applications. *Crit Rev Biotechnol.* 2017;37(2):163-176.
12. Bhagwat N, Dulmage K, Pletcher CHJ, Wang L, Demuth W, Sen M, Balli1 D, Yee SS, Silin Sa, Tong F, Yu L, Moore JS,

- Stanger BZ, Dixon EP, Carpenter EL. An integrated flow cytometry based platform for isolation and molecular characterization of circulating tumor single cells and clusters. *Sci Rep.* 2018;8(1):5035.
13. Handoo A, Dadu T. Flow cytometry in pediatric malignancies. *Indian Pediatrics.* 2018;55(1):55-62.
14. Chernysheva O, Markina I, Demidov L, Kupryshina N, Chulkova S, Palladina A, Antipova A, Tupitsyn N. Bone marrow involvement in melanoma. Potentials for detection of disseminated tumor cells and characterization of their subsets by flow cytometry. *Cells.* 2019; 8(6):627.

---

© 2019 Reis et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*  
*The peer review history for this paper can be accessed here:*  
<http://www.sdiarticle3.com/review-history/50866>